UNIVERSITI TEKNOLOGI MARA

OPTIMIZATION AND EVALUATION OF SARCOCYSTOSIS THERMOSTABLE PCR DETECTION KIT

NURUL FATHIYAH BINTI ZAIPUL ANUAR

MSc

April 2020

AUTHOR'S DECLARATION

I declare that the work in this thesis was carried out in accordance with the regulations of Universiti Teknologi MARA. It is original and is the results of my own work, unless otherwise indicated or acknowledged as referenced work. This thesis has not been submitted to any other academic institution or non-academic institution for any degree or qualification.

I, hereby, acknowledge that I have been supplied with the Academic Rules and Regulations for Post Graduate, Universiti Teknologi MARA, regulating the conduct of my study and research.

| Name of Student | : | Nurul Fathiyah Binti Zaipul Anuar |
|------------------|---|--|
| Student I.D. No. | : | 2016912491 |
| Programme | : | Master of Science (Medicine) – MD780 |
| Faculty | : | Medicine |
| Thesis Title | : | Optimization and Evaluation of Sarcocystosis Thermostable PCR Detection Kit |

| Signature of Student | : | |
|----------------------|---|------------|
| Date | : | April 2020 |

ABSTRACT

Sarcocystis spp. is a parasite that can cause sarcocystosis in human and animals. They are intracellular protozoan parasite that can be found mostly in cattle, sheep and goats. Diagnosis of Sarcocystis spp. is mainly carried by histological investigation or serological methods. Histological examination has limitations, as it depends on the site of collection and load of parasitic infection. On the other hand, serological diagnosis of active infection is also unreliable because activation is not always accompanied by changes in antibody levels. Molecular-based techniques are very efficient, as detection of large sample size can be carried out with enhanced specificity and sensitivity. Hence the most reliable method would be PCR technique. However, the conventional PCR requires trained personnel and cold chain transportation/storage. With the knowledge of the disease caused by the organism, its prevalence and geographical distribution, there is a need to develop and improvise the existing molecular tools which can be handy for early detection of *Sarcocystis* in rural areas, where facilities and molecular biologist might not be available, whilst reducing sample collection to processing time. Thermostable premix to detect Sarcocystis spp. was developed to make conventional PCR more user-friendly. The aim of this study is to develop a sarcocystosis thermostable PCR detection kit and evaluate this kit in terms of the sensitivity and specificity, stability, limit of detection (LOD), and repeatability. PCR amplification was carried out to detect the D2 region in conserved regions of 18S rRNA in Sarcocystis spp. A pair of primers to detect KMT1 gene in Pasteurella multocida was used as internal control. Sarcocystis capranis was used as positive control for this kit. The premix contents were lyophilized in the presence of enzyme stabilizer to ensure the *Taq* polymerase work efficiently at room temperature. A total number of 48 samples were used to test the specificity and sensitivity while the stability of this PCR kit was evaluated using Q10 accelerated aging method. The sensitivity and the specificity of the developed thermostable PCR kit was 100%. The Q10 accelerated ageing method at 37°C was found to be about 2.4 month. The limit of detection (LOD) was evaluated using two-fold dilution DNA of Sarcocystis capranis. The limit of detection is 0.547 ng/µL. In the repeatability test, different samples with various concentrations of DNA samples were used. The repeatability study was used to observe the consistency of the PCR results from thermostable sarcocystosis premix based on the intensity of the bands. The percentage of coefficient of variance (COV) ranged from 1.00 % to 3.22 %. From this result, it is evident that this thermostable PCR kit has a good range of repeatability. The developed sarcocystosis thermostable PCR detection kit is very efficient in detection of Sarcocystis spp. with high sensitivity and specificity with an advantage of being a cold chain free PCR system.

ACKNOWLEDGEMENT

Upon completion of this project, first and foremost, I would like to thank my supervisor, Associate Professor Dr. Jamal Hussaini for the valuable guidance and advice. He inspired me greatly to work in this project and his willingness to spend his time for me had motivated me to do well for my master study. Thanks for his exemplary guidance, monitoring and constant encouragement throughout the course of this thesis. The blessing, help and guidance given by him from time to time shall carry me a long way in the journey of life on which I am about to board.

A special thanks to my co-supervisors, Dr. Navindra Kumari Palanisamy and Professor Dr. Ariza Adnan that also help in my journey. Apart from that, I would like to express a deep sense of gratitude to laboratory staffs of Faculty of Medicine, Centre of the Pathology Diagnostic and Research Laboratories (CPDRL) and Institute of Medical Molecular Biotechnology (IMMB) for their guidance during the experimental work. Besides that, I would like to thank Universiti Teknologi MARA (UiTM) for providing me with facilities to complete this study. An honourable mention goes to my parents, Mr. Zaipul Anuar Abdul Hamid and Mdm. Rokiah Ismail who always gave me motivation. Last but not least, I would like to thank to my fellow friends for their understanding and support in completing this study. Without their help of the particular people that mentioned above, I would face many difficulties while doing this.

TABLE OF CONTENTS

| CONFIRMATION BY PANEL OF EXAMINERS | ii |
|------------------------------------|------|
| AUTHOR'S DECLARATION | iii |
| ABSTRACT | ii |
| ACKNOWLEDGEMENT | iii |
| TABLE OF CONTENTS | iv |
| LIST OF TABLES | viii |
| LIST OF FIGURES | ix |
| LIST OF SYMBOLS | X |
| LIST OF ABBREVIATIONS | xi |
| | |
| | |

| CHA | PTER | ONE: INTRODUCTION | 1 | | | |
|-----|--------------|--|----|--|--|--|
| 1.1 | Resea | Research Background | | | | |
| 1.2 | Proble | em Statement | 4 | | | |
| 1.3 | Objec | tives | 5 | | | |
| 1.4 | Scope | 5 | | | | |
| 1.5 | Signit | 5 | | | | |
| 1.6 | 6 Hypothesis | | | | | |
| | | | | | | |
| CHA | PTER | TWO: LITERATURE REVIEW | 7 | | | |
| 2.1 | Introd | luction | 7 | | | |
| | 2.1.1 | History of Sarcocystis spp. | 7 | | | |
| 2.2 | Struct | cure and Life Cycle of Sarcocystis spp. | 7 | | | |
| | 2.2.1 | Distribution/ Occurrence of Sarcocysts in Animal | 7 | | | |
| | 2.2.2 | Structure of Sarcocysts | 8 | | | |
| | 2.2.3 | Life Cycles of Sarcocystis spp. | 10 | | | |
| | | 2.2.3.1 Definitive Host | 10 | | | |
| | | 2.2.3.1 Intermediate Host | 11 | | | |