

UNIVERSITI TEKNOLOGI MARA

**DEVELOPMENT OF PCR-BASED
DETECTION METHOD FOR FRUIT
BUNCH ROT (FBR) DISEASE IN OIL
PALM**

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MSc

October 2020

AUTHOR'S DECLARATION

I declare that the work in this dissertation was carried out in accordance with the regulations of Universiti Teknologi MARA. It is original and is the results of my own work, unless otherwise indicated or acknowledged as referenced work. This thesis has not been submitted to any other academic institution or non-academic institution for any degree or qualification.

I, hereby, acknowledge that I have been supplied with the Academic Rules and Regulations for Post Graduate, Universiti Teknologi MARA, regulating the conduct of my study and research.


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ABSTRACT

Fruit bunch rot (FBR) disease is considered as a new potential emerging disease of oil palm in Malaysia. The occurrence of this disease has been reported in Perak and Selangor states since 2012. The causal pathogen was identified as *Marasmius palmivorus* Sharples (1928) is known to be as saprophytic apparently become parasitic after it has attained a certain inoculum level of infection. The current study aimed to develop molecular polymerase chain reaction (PCR) detection method for FBR disease in oil palm. The use of molecular diagnostic assay offers specificity and rapid diagnostic for an early disease detection in plantations. Therefore, a total of 39 samples were collected from Perak and Selangor states by random sampling in the oil palm plantations. Basidiocarps and rhizomorphs on diseased oil palm were collected prior surface sterile and 8 pure isolates of *M. palmivorus* were successfully isolated. The identity was confirmed by morphological and molecular identification. An artificial inoculation of *M. palmivorus* on detached oil palm fruit using *in vitro* assay showed positive infection. Ten species-specific primers were designed to amplify the Internal Transcribed Spacer (ITS) of ribosomal gene cluster (rDNA) of *M. palmivorus*. Primers designed in this study were screened for efficiency and specificity before optimized using Gradient PCR for optimum annealing temperature (59 – 64°C) for PCR amplification. Validation assays performed by cross reaction with 34 different fungi species includes 66 samples from different sources. The results obtained suggests that the new designed primer could be employed to directly analyse sample from the field with the aim of detecting the presence of the pathogen in diseased fruits. Molecular analyses suggested that primer pair MP02 was able to amplify *M. palmivorus* at 64 °C annealing temperature in 30 cycles of PCR programme. This report for the first time that species specific primer MP02 has revealed to be useful primer in PCR amplification for the detection of *M. palmivorus* rDNA.

ACKNOWLEDGEMENT

Firstly, I wish to thank God for giving me the opportunity to embark on my master and for completing this long and challenging journey successfully. My gratitude and thanks go to my main supervisor Professor Madya Dr Hasmah Mohidin for her continuous guide, motivation and endless supports on every path I took up myself until this level. I am deeply grateful for what she read some lines of a very incipient research proposal and believed enough with that to accept becoming my supervisor.

Secondly, my appreciation goes to my co-supervisors from MPOB, Director of Biological Research Division, Dr Idris Abu Seman and group leader of Emerging Exotic Biosecurity Disease (EEBD), Dr Maizatul Suriza Mohamed, for their earnest aid and had provided me the facilities and technical assistance throughout the experiment conducted. I would always be indebted to all MPOB staffs (Nor Azmira, Nurzarazilla, Jamil, Rosmidi, Muhd Fazly, Madihah, Nor Shazwana, Intan Nur Ainni, Nur Diyana, Angel Lee, Nur Atiqah, Nur Rasyeda, Dr Salwa, Nur Hailini and Shariffah) for their encouragement and more over for their timely support and precious guidance throughout the laboratory works till the completion of my journey.

Thirdly, I am thankful to and fortunate enough to get constant encouragement, supports and guidance from all FPA lecturers and staffs (Sulaiman Man, Sahmsiah Sahmat, Dr Hendry Joseph, Dr Alex Korom, Hazmi Awang Damit, Rozlianah Fitri, Dr Viduriati, Dayang Shurtini, Dr Siti Hajar, Dr Zaiton Sapak, Norshafizah and Meor Ahmad).

Special thanks are also due to my Master colleagues, namely Muhammad Shahrezaa, Noor Albannia Natasya, Elvera Arene, Ifrah, Mazni, Syazwani, Ahmad Hamdi, Siti Raihana, Shahrizan and Nurazmiuddin, with whom I have shared moments of deep anxiety but also of big excitements. Thank you for always being there for me. I am also very grateful to Steffeny Martina Harley for her amazing sense of respect when helping me with the editing of the thesis.

Finally, this thesis is dedicated to my very dear mother, Sarimah Jamullah and the loving memory of my late father, Jaabi Mohd Suhudi may Allah bless him and grant him a place in heaven, for their vision, trust, determination and financial supports in educate me. This piece of victory is dedicated to both of you. Alhamdulillah.

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