

Potential of Malaysian Cherry Leaves (*Muntingia calabura*) as an Antioxidant Agent

Noor Hidayah Pungot¹*, Nurul Auni Zainal Abidin²*, Nur Syafiqah Atikah Nazaharuddin²

¹Faculty of Applied Sciences, Universiti Teknologi MARA (UiTM), 40450 Shah Alam, Selangor, Malaysia
²Faculty of Applied Sciences, Universiti Teknologi MARA (UiTM), Cawangan Negeri Sembilan, Kampus Kuala Pilah, Pekan Parit Tinggi, 72000 Kuala Pilah, Negeri Sembilan, Malaysia.

Corresponding authors: noorhidayah977@uitm.edu.my and nurulauni@uitm.edu.my Received: 10 April 2020; Accepted: 11 May 2020; Published: 1 June 2020

ABSTRACT

Muntingia calabura has a high phytochemical content, especially the phenolic group that can act as antioxidant. In Malaysia country, this *M. calabura* also known as 'kerukup siam' or 'Ceri Kampung' and it belongs to *Muntingiaceae* family. This research was conducted to determine the potential of antioxidant activity application of cherry leaves (*M. calabura*) from various solvent extracts (methanol, ethyl acetate, and n-hexane). The phytochemical contents were screening by using the established standard procedure. Total phenolic content (TPC) was determined according to the Folin-Ciocalteau colorimetric method, while the antioxidant activity was carried out using 2,2-diphenyl-1-picryhydrazyl (DPPH) radical scavenging assay. Phytochemical screening on the leaves part methanolic extracts revealed that the presence of various biochemicals like flavonoids, phenols, steroids, triterpenes, tannins, reducing sugars, and saponins except the alkaloids. Among the three extracts, the methanol leaf extract gave the highest content of phenolics (8.20 mg GAE/g extract). Analyses of antioxidant activity with IC₅₀ value of 167.70 µg/mL. The present study confirms that the presence of various phytochemicals which shows good antioxidant activity of *M. calabura* leaves. Therefore, it has the potential as a therapeutic antioxidant agent and can be used in cosmeceutical and food products.

Keywords: Muntingia calabura; Ceri kampung; Total phenolic content; Antioxidant

INTRODUCTION

Muntingia calabura is one of the many species from the family *Muntingiaceae* used in folk medicine throughout the world. It is commonly known as the Jamaican cherry in Brazil and 'Kerukup Siam' in Asian region, including Malaysia. Various parts of *M. calabura* such as leaves, barks, flowers, and roots have been employed as a treatment for traditional medical, prevention, rehabilitation and health promotion [1].



Mostly, plants are producing a variety of secondary metabolites or bioactive compounds such as flavonoids [2][3], phenolics [4], tannins [5], quinones [6], glycosides [7] and terpenes [8] that are used as herbal medicine. The scientific evaluations on *M. calabura* have been revealed several pharmacological activities possessed by the plant. *M. calabura* leaves have been reported to exhibit significant anti-inflammatory, antipyretic [9], antinociception [10], antitumor [11], antiproliferative, antioxidant [9], and antibacterial [11] activities. Flavonoids, saponins, tannins, triterpenes, and steroids have been detected also in the leaves of *M. calabura* [3]. Several types of flavonoids have been isolated and identified from the leaves, roots, and stem barks of *M. calabura* [12].

Natural antioxidants have been used also in the cosmetic industries including a great number of substances and extracts obtained from a variety of plants, grains and fruits, either by reducing the skin oxidative stress or protecting the skin from oxidative degradation [13]. Plants that are efficacious as antioxidants are plants that contain carotenoids and polyphenols, especially flavonoids which can be formulated as natural antioxidants in oral dosage forms such as vitamins and topicals for skin care products. Therefore, the aim of this study was to screen the phytochemical present in leaf extracts with three types of solvent polarities using maceration method, to determine the antioxidant activity and total phenolic content of *M. calabura*.

EXPERIMENTAL

Materials and Instruments

The list of solvents used are acetone, methanol, ethyl acetate, and n-hexane were purchased from (HmBG chemicals) while the list of chemicals used are gallic acid, Folin-Ciocalteu reagent were purchased from (R&M Chemical Supplier), and 2,2-diphenyl-1-picrylhydrazyl (DPPH) was purchased from (QReC). Instrument used is T80/T80+ UV-Visible spectrometer in the range 500-700 nm.

Plant material and preparation of extracts

M. calabura leaves were collected in August 2018 from the Alor Gajah, Melaka. The fresh leaves were airdrying for about two weeks at room temperature and grinded into a fine powder. The extracts were prepared by maceration of 100 grams of the powdered sample in methanol, ethyl acetate, and n-hexane (800 mL of each), by using orbital shaker for 72 hrs at room temperature. Then, the extracts were filtered, and concentrated via rotary evaporation (IKA HB 10) at 40 °C. Crude extracts were stored in sealed containers until further analysis [14].

Phytochemical Screening

The leave extracts from of *M. calabura* were subjected to phytochemical screening to test the presence of tannins [15], flavonoids [16], alkaloids, phenols [17], steroids, triterpenes, saponins, and reducing sugar [18] following the standardized methods [19].



Determination of Total Phenolic Content (TPC)

Total phenolic content (TPC) of methanol, ethyl acetate and n-hexane extracts of *M. calabura* leave was carried out by using modified Folin-Ciocalteau colorimetric method [20]. Stock solution of gallic acid was prepared by dissolving 0.01 g of gallic acid and marked up in 100 mL of volumetric flask with distilled water. The mixture was left in the dark for 1 hour. The absorbance of the samples was measured at 760 nm using a UV-Vis Spectrometry. Results were expressed as mg gallic acid equivalent (GAE)/g plant extract. All the experiments were carried out in triplicate. The average absorbance values obtained at different concentrations of gallic acid were used to plot the calibration curve.

Antioxidant assay

The DPPH radical scavenging activity of *M. calabura* has been tested by following the reported method with modifications [21]. The sample in methanol (0.2 mL) with concentrations ranging from 7.81 until 1000 μ g/mL and were mixed with the DPPH solution (3.8 mL). The reaction mixture was then left to stand in the dark for 30 min at room temperature. The absorbance of the reaction mixture was record at 517 nm. Ascorbic acid was applied as a standard antioxidant, while DPPH solution was used as DPPH blank. The percentage inhibition DPPH was calculated using the following Equation 1:

% Inhibition = $[(A_{DPPH blank} - [A_{sample} - A_{blank sample}]) / A_{DPPH blank}] \times 100\%$ (1)

All tests were performed in triplicates and were expressed as mean \pm standard deviation. The graph of percentage inhibition was plotted against the concentration (in µg/mL) to determine the concentration of *M. calabura* extract required to scavenge 50% of DPPH free radicals. The results were reported as IC₅₀ values (in µg/mL).

RESULTS AND DISCUSSION

Phytochemical analysis

Freshly prepared extracts were subjected to a preliminary phytochemical screening for various constituents. The results of the phytochemical analysis of the different extracts (n-hexane, ethyl acetate, and methanol) of cherry leaf have shown a remarkable variation, are presented in Table 1. The methanolic extracts showed presence of secondary metabolite classes due to high quantity of flavonoids, phenols, steroids, triterpenes, tannins, reducing sugars, and saponins with absence of alkaloids.

On the other hand, ethyl acetate and n-hexane extract revealed the flavonoids, phenols and steroids. These constituents have ability to be hydrogen donor makes it suitable to acts as antioxidants [22]. Meanwhile, the three of solvent extraction was absent in the alkaloids test for *M. calabura*. The present study regarding the qualitative analysis of the selected medicinal plants is in agreement with the previous findings by the various researchers [23-27].



Leaf Extracts	Constituents							
	Alkaloids	Flavonoids	Phenols	Steroids	Tannins	Saponins	Triterpenes	Reducing sugars
n-hexane	-	+	+	+	-	-	+	-
E. Acetate	-	++	++	+	-	-	-	-
Methanol	-	+++	+++	++	++	++	++	+

Table 1: Phytochemical profiles of Cherry leaf (*M.calabura*) extracts

- = absent; + = less present; ++ = moderate; +++ = high [28]

Based on the above discussion, the phytochemicals present in the crude extracts of *M. calabura* leaf can serve as a valuable source of information and provide appropriate standards to establish a base for identification and elucidation of the different types of bioactive chemicals.

Quantitative Total Phenolic Content and DPPH Radical Scavenging Activity

Total phenolic content and DPPH radical scavenging activity of n-hexane, ethyl acetate, and methanol extracts of the leaves of *M. calabura* are presented in Table 2. Among the extracts, the methanol extract was displayed the highest total phenolic content (8.20 mg GAE/g) and the lowest IC₅₀ value (167.70 μ g/mL) as compared to the ethyl acetate and n-hexane extract. In this study the DPPH method was used to obtain IC₅₀ values from a plant extract.

 IC_{50} is the concentration of the sample to inhibit 50% of free radicals. Experimentally, the purple color of DPPH solution decolorized into yellow color as hydrogen from the antioxidant source was accepted. The lower the IC₅₀ value, the higher the antioxidant properties of the extracts sample and vice versa [29]. Based on the results of the study of IC₅₀, leaves methanol extract of *M. calabura* is very strong antioxidant due to the lower value of DPPH, while for IC₅₀ of leaves n-hexane extract is slightly higher due to the lower of total phenolic content.

Table 2: Total phenolic compounds (TPC) and antioxidant activity (DPPH) of different phenolics fractions of Cherry leaf (*M. calabura*) extracts

	Solvents						
Assay —	n-hexane	Ethyl Acetate	Methanol				
TPC (mg GAE/g extract) ^a	2.80±0.0	4.42±0.0	8.20±0.0				
DPPH (IC ₅₀ , µg/mL) ^a	408.80±0.5	404.03±0.7	167.70±0.6				

^aData represent mean \pm standard deviation of three replicate experiments;

Positive control (ascorbic acid = $10.68 \pm 0.9 \ \mu g/mL$)



It was reported that the IC₅₀ values obtained from *M. calabura* leaves extracts were: 496.18±4.56 µg/mL for petroleum ether extract, 107.99±6.24 µg/mL for chloroform extract, 79.96±0.91µg/mL for ethanol extract, and 97.638±2.06 µg/mL for aqueous extract in comparison to ascorbic acid 40.43±3.95 µg/mL [30]. However, their IC50 values were higher than that of ascorbic acid (10.68 µg/mL). It can be seen that the methanol extract of *M. calabura* has the strongest antioxidant activity because it has a very high content of phenolic compounds. The radical scavenging activity of the extracts were in the order of polarity of solvent methanol > ethyl acetate > n-hexane.

Also, the result are agreeing with the previous study who are reported that the methanol extract of *M*. *calabura* was shown the significant antioxidant activities, due to high containing of phenolic compounds [31]. From the phytochemical screening, the flavonoids and phenolics compound had the strongest presence in methanol extracts of *M. calabura* as shown in Table 1. These compounds were give the inhibition against the free radical for antioxidants assay. Therefore, these results indicated that the antioxidant activity of extracts was well correlated with TPC [32].

CONCLUSIONS

The phytochemical analysis showed that the leaves extracts of *Muntingia calabura* contain flavonoids, phenols, steroids, triterpenes, tannins, reducing sugars, and saponins. The significance TPC value and the potent antioxidant activity (DPPH) of the leaf methanol extract indicated that this plant could be beneficial as a source of natural antioxidant. From these findings, it is suggested that the methanol extract can be further subjected for the isolation and characterization of phytochemicals that have antioxidant effect.

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REFERENCES

- [1] Nshimo, C.M., Pezzuto, J.M., Kinghorn, A.D., and Farnsworth N.R. Cytotoxic constituents of *Muntingia calabura* leaves and stems collected in Thailand. *Int. J. Pharmacol*, 31, 77-81 (1993).
- [2] Pahari, B., Chakraborty, S., Chaudhuri, S., Sengupta, B., and Sengupta, P.K. Binding and antioxidant properties of therapeutically important plant flavonoids in biomembranes: insights from spectroscopic and quantum chemical studies. *Chem Phys Lipids*, 165 (4), 488-496, (2012).
- [3] Sufian, A.S., Ramasamy, K., Ahmat, N., Zakaria, Z.A., and Mohd. Yusof, M.I. Isolation and identification of antibacterial and cytotoxic compounds from the leaves of *Muntingia calabura*. *J. Ethnopharmacol*, 146, 198-204, (2013).
- [4] Dai, J., and Mumper, R.J. Plant phenolics: extraction, analysis and their antioxidant and anticancer properties. *Molecules*, 15 (10), 7313-7352, (2010).
- [5] Wong, I.L., Chan, K.F., Chen, Y.F., Lun, Z.R., Chan, T.H., and Chow, L.M. *In vitro* and *in vivo* efficacy of novel flavonoid dimers against cutaneous leishmaniasis. *Antimicrob Agents Chemother*, 58 (6), 3379-3388, (2014).



- [6] Basavegowda, N., Sobczak-Kupiec, A., Malina, D., Yathirajan, H.S., Keerthi, V.R., Chandrashekar, N.P., Liny, P. Plant mediated synthesis of gold nanoparticles using fruit extracts of Ananas comosus (L.)(Pineapple) and evaluation of biological activities. *Adv. Mat. Let*, 4 (5), 332-337(2013).
- [7] Mithöfer, A. and Boland, W. Plant defense against herbivores: Chemical aspects. *Annu Rev Plant Biol*, 63, 431-450, (2012).
- [8] Buhian, W., P., C., Rubio, R., O., Valle, D., L., and Martin-Puzon, J., J. Bioactive metabolite profiles and antimicrobial activity of ethanolic extracts from Muntingia calabura L. leaves and stems. *Asian Pacific Journal of Tropical Biomedicine*, 6(8), 682–685, (2016).
- [9] Zakaria, Z.A., Kumar, G.H., Mohd Zaid, S.N.H., Abdul Ghani, M., Hassan, M.H., Mohd Nor Hazalin, N.A., Khamis, M.M., Devi, R.G. Analgesic and antipyretic actions of *Muntingia calabura* leaves chloroform extract in animal models. *Orient. Pharm. Exp. Med.*, 7, 34-40, (2007).
- [10] Zakaria, Z.A., Mohd Nor Hazalin, N.A., Mohd Zaid, S.N.H., Abdul Ghani, M., Hassan, M.H., Gopalan, H.K., and Sulaiman M.R. Antinociceptive, anti-inflammatory and antipyretic effects of *Muntingia calabura* aqueous extract in animal models. *J. Nat. Med.*, 61, 443-448, (2007).
- [11] Su, B.N., Jung Park, E., Vigo, J.S., Graham, J.G., Cabieses, F., Fong, H.H., Pezzuto, J.M., and Kinghorn, A.D. Activity-guided isolation of the chemical constituents of *Muntingia calabura* using a quinone reductase induction assay. *Phytochemistry*, 63, 335-341, (2003).
- [12] Mahmood, N.D., Nasir, N.L., Rofiee, M.S., Tohid, S.F.M., Ching, S.M., Teh, L.K., et al. *Muntingia calabura*: A review of its traditional uses, chemical properties, and pharmacological observations. *Pharm Biol*, 52 (12), 1598-1623, (2014).
- [13] Zakaria, Z.A., Sufian, A.S., Ramasamy, K., Ahmat, N., Sulaiman, M.R., Arifah, A.K., et al. In vitro antimicrobial activity of *Muntingia calabura* extracts and fractions. *Afr J Microbiol Res*, 4 (4), 304-308, (2010).
- [14] Buhian, W. P. C., Rubio, R. O., and Martin-Puzon, J. J. Chromatographic fingerprinting and free-radical scavenging activity of ethanol extracts of *Muntingia calabura* L. leaves and stems. *Asian Pacific J. of Tropical Biomedicine*, 7(2), 139-143, (2017).
- [15] Preeti, S., Gaurava, S. Pharmacological and phytochemical screening of *Desmodium gangeticum* and *Moringa oleifera. Research Journal of Chemistry and Environment*, 22 (5), 6-10, (2018).
- [16] Shanmugam, U., K., Nagajaran, K., Natarajan, K., Thiraviam, P., P., Thirayagarajan, B., and Sekkizhar, G. Preliminary phytochemical analysis of sidha formulation- parangipattai kudineer. *European Journal of Biomedical and Pharmaceutical Sciences*, 49 (1), 526-530, (2017).
- [17] Kadirvelmurugan, V., Tamilvannan, M., Arulkumar, M., Senthilmurugan, V., Thangaraju, K., Dhamotharan, R., and Ravikumar, S. Phytochemical, HPTLC finger print analysis and antimicrobial activity of ethyl acetate extract of *Decalepis hamiltonii* (Wight & Arn.). *Journal of Academia and Industrial Research*, 5 (9), 126-131, (2017).
- [18] Ukwubile, C., A., Samagoro, C., T., and Nuhu, A. Preliminary phytochemical screening and acute toxicity determination of *Camellia sinensis L. (Theaceae)* leaf methanol extract in swiss albino mice. *International Journal of Biological Sciences and Research*, 1 (1), 1-17, (2018).
- [19] Costa, R., and Santos, L. Delivery systems for cosmetics From manufacturing to the skin
- of natural antioxidants. Powder Technology. 322: 402-416 (2017).
- [20] Singleton, V.L., and Rossi Jr, J.A. Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *American Journal of Enology and Viticulture*, 16,144-158, (1965).
- [21] Blois, M.S. Antioxidant determinations by the use of a stable free radical. *Nature*, 181, 1199-1200, (1958).
- [22] Singh, R, Iye, S, Prasad, S, Deshmukh, N., Gupta, U., Zanje, A., Patil, S., and Joshi, S. Phytochemical Analysis of *Muntingia calabura* extracts possessing anti-microbial and anti-fouling activities. *International Journal of Pharmacognosy and Phytochemical Research*, 9(6), 826–832, (2017).
- [23] Singh, R, Iye, S, Prasad, S, Deshmukh, N., Gupta, U., Zanje, A., Patil, S., and Joshi, S. Phytochemical Analysis of *Muntingia calabura* extracts possessing anti-microbial and anti-fouling activities. *International Journal of Pharmacognosy and Phytochemical Research*, 9(6), 826–832, (2017).



- [24] More, S., Upadhye, M., Lohakare, A., and Jagtap, S. Comparative quantification of flavonoid content and antioxidant potential of indigenous medicinal plants. *Journal of Pharmacognosy and Phytochemistry*, 7 (1), 343-345, (2018).
- [25] Fariestha, G., A., K., Andayani, S., Yanuhar, U. Analysis of the secondary metabolite of kersen leaf extracts (*Muntingia calabura L.*) and its potential as anti-bacteria to inhibit *Aeromonas hydrophila*. *Research Journal of Life Science*, 5 (2), 121-127, (2018).
- [26] Desrini, S., and Ghiffary, H., M. Comparison of antibacterial activity of Talok (*Muntingia calabura L*) leaves ethanolic and n-hexane extracts on Propionibacterium acnes. *AIP Conference Proceedings*, 1954 (1), 030002, (2018).
- [27] Vasanth, M., Selvaraju, S., Muralidharan, R., and Rajarajan, R. Antimicrobial activity of *Muntingia calabura L. World Journal of Pharmaceutical Research*, 6 (11), 663-667, (2017).
- [28] Yemineni, M., Kancharlapalli, V. R., & Pn, S. Preliminary phytochemical investigation and evaluation of hypoglycemic activity of methanolic extract of *Muntingia Calabura L* stem bark against normal and streptozotocin-induced diabetes. *Asian Journal of Pharmceutical Clinical Research*, 12 (6), 137-142, (2019).
- [29] Yudden, N. k. M., Nordin, N. N., Harun, A., Rosli N. H., Aziz, N. A, and Daud, S. Comparative Study of Antioxidant Activity of Stem and Leaves of *Entada Spiralis* and Their Antibacterial Properties Against *Erwinia Chrysanthemi. Science Letter*, Vol. 13 (2), 38-47, (2019).
- [30] Aruna, S., M., Bodke, Y., D., and Chandrashekar, A. Antioxidant and in vivo anti-hyperglycemic activity of *Muntingia calabura* leaves extracts. *Der Pharmacia Lettre*, 5(3), 427-435, (2013).
- [31] Zakaria, Z. A., Mahmood, N. D., Omar, M. H., Taher, M., & Basir, R. Methanol extract of *Muntingia calabura* leaves attenuates CCl4-induced liver injury: possible synergistic action of flavonoids and volatile bioactive compounds on endogenous defence system. *Pharmaceutical Biology*, 57 (1), 335–344, (2019).
- [32] Özbek, H. N., Halahlih, F., Göğüş, F., Koçak Yanık, D., & Azaizeh, H. Pistachio (*Pistacia vera L.*) hull as a potential source of phenolic compounds: evaluation of ethanol–water binary solvent extraction on antioxidant activity and phenolic content of pistachio hull extracts. *Waste and Biomass Valorization*. 11, 2101–2110 (2020).